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A. M. CLARK, M.A.

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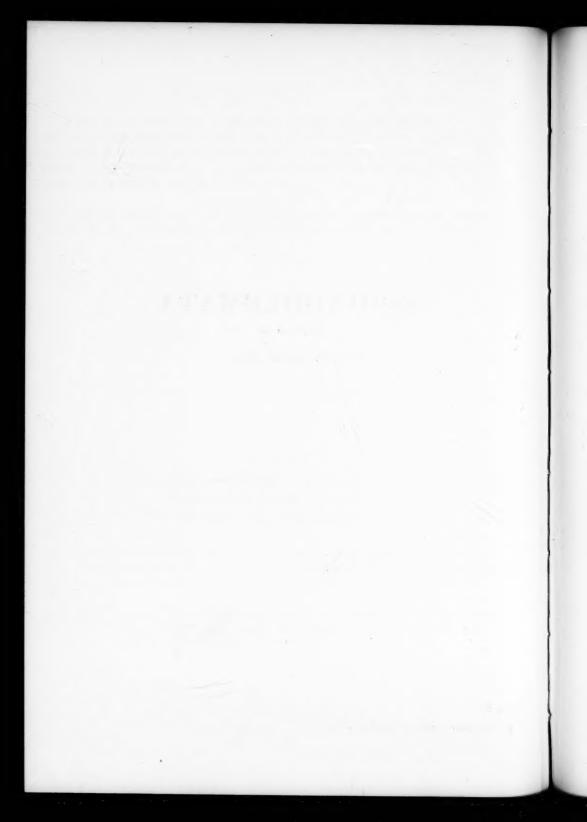
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## II.—SUBJECT INDEX

## GENERAL LITERATURE AND HISTORY

Text-books.—Palaeozoology, Ehrenberg; encyclopedia articles on various major groups, Fell (3); amateur fossil collector's handbook for Texas, U.S., Matthews; biology of marine animals, Nicol; Tertiary stratigraphy, Papp; micro-palaeontology, Pokorný; stratigraphie palaeontology, Termier & Termier.

Bibliography.-Of the Danian and Paleocene, Brotzen.

Nomenclature.—Proposed suppression of Spatagus O. F. Müller, 1776, Melville (1); proposed stabilization of Echinus minutus Buckman, 1845, Melville (2); proposed validation of Encrinus, Spillane; keys to the genera of Ophiuroidea with synonyms, Fell (2).

Collections.—Of the Oceanographic Institute, University of San Pablo, Brazil, asteroids, Bernasconi (1); catalogue of type fossils of the Geological Survey of Canada, Bolton.

Obituary.-Louis Castex (1890-1955), Magne.

#### STRUCTURE

Microstructure.—Sectioning sea urchin spines, Anon. (2). Needham; structures that may be adaptations for cell adhesion, Afzelius; histological studies on digestive system of starfish Henricia, Anderson (2); yolk globules

of oocytes, Bolognari; secretory structures in starfish tube foot, Chaet & Philpott; structure of ground cytoplasm as revealed by electron microscopy, Gross, Philpott & Nass; of tube-feet of Antedon bifida, Nichols; starfish eye-spot, Philpott & Chaet; origin of the nucleolus, Vincent.

Skeleton.—Crystallography of echinoid calcite, Raup; statistical study of holothurian sclerites, Hampton.

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Digestion.—Functions of Tiedemann's pouches in Henricia and other starfishes, Anderson (2); transport of nutrients in a synaptid holothurian, D'Agostino & Farmanfarmaian; of alga Macrocystis by echinoid Strongylocentrolus, Lasker & Boolootian.

Respiration.—Cytoplasmic viscosity in the echinoid egg during suppressed respiration, Butros (1); in mature echinoid eggs, Gonse; release of respiratory control by DNP. Immers & Runnström.

Growth.-Of echinoid test, Raup, Swan (1).

Pigments.—Of fossil crinoid, Blumer; naphthoquinone derivatives, Kuroda & Okajima; echinochrome A from *Diadema* spines, Nishibori; from dorsal skin and eyespots of Asterias, Rockstein and others.

Regeneration.—Of the cardiac stomach of Asterias, Anderson (1).

Muscular activity.—The motility-inducing factor of flagellum and the relaxing factor of muscle, Kinoshita; longitudinal muscles of *Holothuria grisea*, Sawaya & Ancona Lopez.

Bio-chemistry.—Determination of acetylcholine content from electric organ extract of the ray Narcine tested on longitudinal muscle of Holothuria, Ambache & Sawaya; estradiol- $17\beta$  and progesterone in starfish ovaries, Botticelli, Hisaw & Wotiz; sterol biosynthesis, Pagerlund & Idler; effect of sugars on differentiation of larvae, Hörstadius (2); mucoprotein in holothurians, Motohiro; biochemical approach to cell morphology, Vincent.

Toxicity.—Toxic factor in scalded starfish, Chaet & Cohen; the structure of holothurin, Chanley and others; saponin, the toxic principle of starfish, Hashimoto & Yasumoto; poison effects of echinoderms, Albahary & Budker; toxicity: of holothurin from Actinopyga agassizi, Friess and others, Nigrelli & Jakowska, of holothurians, Frey; toxic substance in starfish stomach, Nakazawa; effect of holothurin on echinoid development, Ruggieri & Nigrelli.

## REPRODUCTION AND SEX

Reproductive cycles.—Gonad development in Asterina gibbosa, Delavault (1) and (2); cycle in Strongylocentrotus, Puii.

Spawning.—Induced discharge of starfish gametes, Chaet & Musick; temperature range for spawning, Mileikovsky.

Sexuality.—Of Echinaster sepositus, Cognetti & Delavault (2), of Asterina, Delavault (3), of Strongylocentrotus, Fuji.

Sex maturation.—Effect of temperature on gonadal growth, Boolootian; size of echinoids at sex maturity, Fuji.

Sexual dimorphism.—In echinoids, Tahara, Okada & Kobayashi.

Hermaphroditism.—In Atlantic and Mediterranean asteroids, Cognetti & Delavault (1), in Arbacia punctulata, Zimmerman, Zimmerman & Harvey.

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Spermatozoa.—Morphology of starfish sperm., Bernstein & Fehrenbaker; fine structure of the acrosome, Dan.

Fertilization including experimental studies.—Allen; effect of antibodies on fertilization rate, Hultin; fertilization-inhibiting action of dermal secretion, Metz; transfer of potassium across membrane before and after fertilization, Tyler & Monroy.

Cleavage.—Position of first cleavage furrow, Endo; cell division blocked by heavy water, Gross & Spindel; cleavage with nucleus intact, Harvey; acceleration of cleavage by vital staining with neutral red, Kojima; induction of cleavage furrows in eggs, Marsland, Zimmerman & Auclair; effect of tensile stress on cleavage, Rappaport; furrowing in flattened eggs, Soott (1); surface changes during cell division, Scott (2); effect of 3, 3-dimethyl-1-phenyltriazene on cleavage rate, Stevens & Turner; retardation of cleavage by cell extracts, Wolfson; SH groups and retardation of cleavage, Wolfson & Wilbur; induction of furrowing reaction, Zimmerman & Marsland.

Mitosis.—In hybrid echinoid embryos, Baltzer & Chen; multiplicity of mitotic centres, Mazia, Harris & Bibring; changes in sensitivity to ultraviolet-induced mitotic delay during cell division cycle, Rustad; nuclear-cytoplasmic relations, Rustad & Rustad; mitotic variations and the rate of O<sub>3</sub>-consumption, Zeuthen; physico-chemical analysis of the isolated mitotic apparatus, Zimmerman.

Gastrulation.—Cellular mechanisms in morphogenesis of gastrula, Gustafson & Kinnander; cellular basis of early gastrulation, Kinnander & Gustafson.

Larvae.—Abnormally symmetrical plutei produced by treatment with colchicine, Csihak (1); larvae of Holothuria glaberrima and Echinometra lucunter, Lewis; plutei, Ubisch.

Experimental embryology.—Stimulation of cleavage by DNA fractions, Butros (2); effect of colchicine in producing abnormal plutei, Cnihak (1); vegetalization by tyrosine, Fudge; cytoplasmic structure in centrifuged egg, Gross, Philpott & Nass; effect of sugars on differentiation, Hörstadius (1); effect of \$\frac{2}{3}\$ and of X-irradiation on echinoid eggs, Hsiao, Fujii & Daniel; cytoplasmic components of eggs stratified by centrifuging, Immers (2); release of respiratory control by DNP, Immers & Runnström; effect of: fatty acids on respiration in eggs, Ishihara, of partial protein extraction on egg structure, Kane, of FUDR, etc., on embryo, Karnotsky & Basch, of salyrgan, an organic derivative of mercury, on development of echinoid egg, Lallier (1), of 2-desoxy-D-glucose, Lallier (2), of alkaline metal ions on development, Lallier (3), and of dyes with a halogenated triazine group on development, Lallier (4); inhibition of cleavage by ribonuclease, Ledoux & Mets; inhibitor of cytochrome oxidase activity, Maggio & Monroy; effect of irradiation on nuclear function, Heifakh; incorporation of thymidine in developing

eggs, Nigon & Nonnenmacher; effect of crysoidin seawater and of vitamin B<sub>a</sub> on development, Okada & Shimoizumi; inhibited breakdown of cortical granules in development, Osanai; effect of 3:5:3-tri-iodothyronine and its derivatives on development, Roche, André & Bouxin; inhibiting effect of nitrogen mustard on development, Roguski; effects of hyperatmospheric O<sub>z</sub>-concentrations on early cleavage, Rosenbaum & Wittner; animalizing and other effects of holothurin on development, Ruggieri & Nigrelli.

Chemical embryology.—Activity of glucose-6-phosphate dehydrogenase, Backström (1): reduction of blue tetrazolium, Backström (2); pathways of glucose metabolism, Backström, Hultin & Hultin; synthesis of nucleic acids in hybrid embryos, Baltzer & Chen; nucleic acids, Chen; incorporation of labelled aminoacids into proteins of mitochondria, Giudice; protein metabolism of embryos, Hultin & Bergstrand; incorporation of labelled amino-acids, Immers (1); SH and S-S groups during first cleavage, Kawamura and Kawamura & Dan; antigens of sperm. surface, Köhler & Metz; inhibitor of cytochrome oxidase in unfertilized eggs, Maggio, Ajello & Monroy; antigens of sperm. extracts, Mets & Köhler; incorporation of S<sup>25</sup>-methionine during early development, Monroy; glycoprotein of eggs, Monroy & Vittorelli; secretion of mucosubstance in gastrula, Motomura; enzymatic activity of ribonuclease Nair; cytochemical localizations and ultrastructure in the fertilized unsegmented egg, Pasteels, Castiaux & Vandermeerssche; enzyme activity in echinoid eggs, Pimpinelli; 5-deoxyuridic acid, Scarano; DNA synthesis during early development, Scarano & Maggio; enzyme APT in eggs, Yanagisawa; hatching enzyme, Yasumasu.

Metamorphosis.—Of Psammechinus, Czihak (2); of echinoids, Pfeiffer; and phylogeny, Uchida.

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Phylogeny.—"Intermediate" forms among early Palaeozoic echinoderms, Regnéll (1); the evidence of metamorphosis, Uchida.

Variation.—Of fossil echinoid Ilarionia arnaudi, Chabaglian; in sea urchins, Swan (1).

Ontogeny.—Of fossil echinoids, Devriès, of fossil Pelmatozoa, Regnéll (3).

#### ECOLOGY AND HABITS

General Works .- The shores of Mozambique, Kalk.

Palaeoecology.—Of the Upper Denton formation, Texas, Laughbaum.

Feeding.—Reaction of scallops to starfishes, Baird; of Asterias, Burnett; of sand dollar Mellita, Goodbody; of Strongylocestrotus on alga Macrocystis, Lasker & Boolootian; carrion eating of brittle-stars, Magabhushanam & Colman; tube-feet in feeding of Antedon, Nichols; Ophiothrix and Ophiopholis but not Ophiocomina, feeding on phytoplankton, Roushdy & Hansen; of Echinarachnius, Sokolova & Kurnetsov; of starfish

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Plankton.—Qualitative and quantitative studies, Legaré & Maclellan; off Portuguese Guinea, Santos Pinto; of Antarctic, Senő; of Sea of Japan, Vinogradov.

Parasites and Commensals.—Fish Carapus commensals with the holothurian Thyone, Ancona Lopez; holotrich clilates endocommensal in echinoid Strongylocentrotus echinoides, Berger; copepods of genera Cancerilla, Ophioika and Lernacosaccus parasitic on antarctic spp. of Ophiacantha, also Codoba on Ophiura; asocthoracic cirripid Ascothorax on Amphiura spp., Heegard; endoparasitic gastropods in holothurians of the state of Washington, U.S., Tikasingh; copepod Cancerilla tubulata on Amphipholis squamata, fig., Zavodnik (1) and (2).

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Mediterranean and Black Sea.—Sardinia, planktonic larvae, Anichini; Black Sea holothurian, Cherbonnier (1); S. France, holothurians, Cherbonnier (2); S. France,

Costa; Algeria, Dieuzeide; S. France, Kerneis; E. Spain and Balearic Is., Sanchez; Naples, starfish, Tortonese (2); Sea of Marmara and the Bosphorus, Tortonese & Demir; Adriatic, Zavodnik (1).

Tropical Atlantic.—W. Indies, W. Africa, Engel, Croes & Schroevers; Gulf of Mexico, Kornicker and others; Barbados, Lewis; Guiana, starfish, Moore (2).

South Atlantic.—Brazil, asteroids, Bernasconi (1); Brazil, Brito (1) to (5); Brazil, echinoid, Krau; Brazil, all but ophiuroids, Tommasi.

Indo-W. Pacific.—Japan, photographs, mostly of living Echinoderms, Baba; Queensland, Bennett; Chatham Islands, E. of New Zealand, Fell (1); Mozambique, Kalk; Okinawa Is., Okada; Red Sea, Tortonese (1).

East Pacific.—El Salvador and California, Aberson & Engel; S. California, Hartman & Barnard.

North Pacific.—Sakhalin and the Kurile Is., fauna list, Diakonov, Baranova & Saveljeva; Bering Sea, Neuman; Hokkaido, N. Japan, and Kurile Is., echinoids, Utinomi; Sea of Japan, plankton, Vinogradov.

Antarctic.-Plankton, Seno.

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Miocene.—Patagonia, echinoids, Bernasconi (3); Roumania, echinoids, Givulescu & Duşa; Czechoslovakia, Kalabis; Austria, ophiuroids, Küpper; Yugoslavia, Mitrović-Petrović; Papp; Austria scutellids, Schaffer; Japan, echinoids, Shibata; I. of Porto-Santo, near Madeira, Silva; Hungary, Somos & Kókay.

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Eocene.—Britain, Anon. (1); France, Castex; Roumania, Ion; Papp.

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Cretaceous.—Echinoids of Caucasus, Askerov & Mamedzade; Patagonia, echinoids, Bernasconi (3) (Danian) Sweden, echinoids, Brotzen; Colombia, Bürgl; Kent, England, Casey; France, Castex; Texas, Laughbaum; Poland, Lefeld & Radwanski; Yugoslavia, Mitrovió-Petrovió; Greece, Mitsopoulos; N.E. Germany, large asteroids, Müller; Lower Silesia, Radwańska; U.S.S.R., Schmidt & Vereshchaghin; Hungary, Szörényi; Chile, comatulids, Wetzel.

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Ordovician.—Ohio, U.S., edricasteroids, Kesling & Mints; S. Korca, Kobayashi; British Isles and Baltic countries, Regnéll (2).

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Sub-phylum Pelmatozoa, other than Crinoidea, **Fell (3)** vol. 2 pp. 256, 532 8 figs.; vol. 3 p. 649; vol. 4 pp. 209, 412 4 figs.; vol. 5 p. 32; vol. 9 pp. 544, 615.

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† Rhabdotites dorsetensis, Hampton pp. 307-314 pl. i (with synonyms R. divergens, bifidus, tridens and irregularis all of Hodson, Harris & Lawson, 1956.)

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†Anorthopygus orbicularis, Castex pp. 38-40, pl. ii figs. 12, 13.

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Apatopygus recens, Fell (1) p. 72.

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†Arbacia fossil and recent p. 107 with A. incisa p. 108 pl. xxvi figs. 6, 8, 10, Hertlein & Grant.

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†Brisaster fossil and recent pp. 131-132 with B. townsendi woynari subsp. nov. Pliocene S. California, Hertlein & Grant pp. 132-133 pl. xxv fig. 5, pl. xxvi figs. 1-3.

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†Brissopsis (Brissoma) ottnangensis, Givulescu & Duşa pp. 937-938 figs. 5, 6; B. ottnangensis, Kalabis pp. 105-106 fig. 3.

† Brissus Duperieri sp. nov. Oligocene France, Castex pp. 33-34 pl. ii figs. 21, 22.

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† Echinocyamus Schoelleri sp. nov. Oligocene France p. 31, E. Touzini pp. 40-41, Castex; E. pusillus, Anon. (1) pl. xxxi fig. 3.

† Echinodiscus marginalis var. Tenuissima, Castex p. 36.

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† Echinopedina (Stereopedina) ameghinoi, Bernasconi (3) p. 145.

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†Fibularia (Eoscutum) seefriedi, Chabaglian pp. 151-152 pl. i figs. 15-17.

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†Hemicidaris jaisalmerensis sp. nov. Jurassic Rajasthan, India, Sahni & Bhatnagar in Sahni p. 187, Sahni & Bhatnagar pp. 431-433 pl. iv figs. 11-17.

†Hemipneustes (Spatagoides) striato-radiatus, Askerov & Mamedzade pp. 46-47 fig. 2.

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†Pseudooffaster caucasicus, Askerov & Mamedzade pp. 47-49 fig. 3 a-e.

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†Spatangus fossil and recent p. 128, with S. rarus pp. 128-129 pl. xxv figs. 1-3, Hertlein & Grant.

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[Vacant]

### ASTEROIDEA

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Sub-genus Amphispina Nielsen, 1932, of genus Amphiodia, a synonym of Ophiophragmus Lyman, 1865, Fell (2) p. 22.

Amphistigma H. L. Clark, 1938, a provisional synonym of Ophiostigma Lütken, 1856, Fell (2) p. 22.

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Ophiochondrella Verrill, 1899 referred to family Ophiacanthidae, Fell (2) p. 18.

Ophiocoma scolopendrina, Baba p. 146 figs.

Ophiocrasis H. L. Clark, 1911 probably no more than a subgenus of Ophionereis, Fell (2) p. 26.

Ophiocten sericeum, Djakonov (2) p. 298.

Ophiocyclus H. L. Clark, 1939, doubtfully distinct from Amphigyptis Nielsen, 1933 ['32], in family Hemieuryalidae, Fell (2) p. 12.

Ophiodera Verrill, 1899 provisionally valid, Fell (2) p. 14.

Ophioderma longicaudum, Zavodnik (1) p. 13.

Ophiodesmus Ziesenhenne, 1940 valid, Fell (2) p. 26.

Ophiodyscrita H. L. Clark, 1938 probably the juvenile form of Cryptopelta H. L. Clark, 1909, Fell (2) p. 27.

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Ophiomisidium irene, Fell (1) p. 69.

†Ophiomusium cf. gagnebini, Hess (4) pp. 389-396 figs. 1-6, 16-23.

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†Ophiopinna gen. nov. pp. 375-376, with type Geocoma elegans Heller, Hess (3) pp. 347-371 figs. 1-34, pp. i; O.? wolburgi sp. nov. Jurassic Switzerland, Hess (4) pp. 411-420 figs. 24-38.

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Ophiopyren stratium, Djakonov (1) pp. 328-329, 332 fig. 8.

Ophiopyrgoides H. L. Clark, 1939 possibly represents a growth stage of Ophiopyrgus Lyman, 1878, Fell (2) p. 31.

Ophiosemnotes Matsumoto, 1917 a synonym of Ophiophthalmus Matsumoto, 1917, Fell (2) p. 18.

Ophiostegastus Murakami, 1944, a synonym of Ophiocormus H. L. Clark, 1915, Fell (2) p. 28.

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†Ophiothrix? royeri, Hess (4) pp. 396-401 figs. 7, 8.

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†Ophiotrigonum gen. nov. (family uncertain) for type O. oxfordiense sp. nov. Jurassic Switzerland, Hess (4) pp. 401-411 figs. 9-15.

Ophiotriton Döderlein, 1896, probably no more than a subgenus of Ophionereis Lütken, 1859, Fell (2) p. 26.

Ophiotylos Murakami, 1943 very close to Ophiozonoida H. L. Clark, 1915, Fell (2) p. 33.

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Ophiuridae Lyman, 1865 used in place of Ophiolepididae Ljungman, 1867, Fell (2) pp. 1, 28.

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Pectinura gracilis, maculata, Fell (1) p. 69.

Stegophiura nodosa, Djakonov (2) p. 298.

"Stella laevis" Rondelet, 1554 probably an Ophioderma, Fell (2) p. 27.

#### CRINOIDEA

Crinoidea, Fell (3) vol. 1 pp. 566–567 fig.; vol. 2 p. 432 fig.; vol. 3 pp. 547–550 10 figs.; vol. 5 p. 300 fig.; vol. 7 p. 49 fig.

†Actinocrinites nodosus, Pickett p. 88 figs. 1, 2.

†Allagecrinus bassleri, Strimple (1) pp. 116-117 pl. i figs. 18-23.

Antedon mediterranea, Zavodnik (1) p. 8.

†Apographiocrinus typicalis, Strimple (1) pp. 117-120 pl. i figs. 1-11, pl. ii fig. 1.

† Balanocrinus subbasaltiformis, Anon. (1) pl. iv figs. 8,9.

Bathycrinus (Ilycrinus) carpenteri, Djakonov (1) pp. 330-331, 332 fig. 9.

†Calceolispongidae gen. et. sp. nov., Permian W. Australia, Dickins & Thomas pp. 78-79.

Cenocrinus Thomson, 1864, with type species Isis asteria Linnaus, 1767, proposed to go on the Official List of Generic Names in Zoology, Spillane pp. 65-68.

†Encrinus Lamarck, 1801, with type species E. liliiformis Lamarck designated, proposed to go on the Official Lists of Generic and Specific Names in Zoology, Spillane pp. 65-68; E. cassianus pp. 543-544, granulosus pp. 544-545, E. cf. liliformis pp. 545-546, E. monetensis p. 546, E. cf. varians pp. 546-547, Virgili.

†Erisocrinus typus, Strimple (1) pp. 120-121 pl. i figs. 14-17, pl. ii figs. 2-5.

†Ethelocrinus expansus p. 127 pl. ii figs. 8, 10, plattsburgensis p. 126 pl. ii figs. 6, 9, 12, Strimple (1).

†Galateacrinus Allisoni, Strimple (2) pp. 195-196.

Heliometra glacialis, Djakonov (2) p. 300.

†Kallimorphocrinus angulatus, Regnéll (3) fig. 6 p. 780.

† Moscovicrinus bijugus, Ivanova frontispiece.

†Paragassizocrinus pp. 5-8, with P. asymmetricus pp. 15-18 pl. i figs. 1-5, deltoideus pp. 20-22 pl. i figs. 11-17, disculus pp. 23-24 pl. iii figs. 1-3, elongatus pp. 18-19 pl. ii figs. 1, 2, hoodi pp. 22-23 pl. i figs. 9, 10, springeri p. 24 pl. ii figs. 6, 7, turris pp. 19-20 pl. ii figs. 6-8, spp. nov. Carboniferous Oklahoma, U.S., P. mequirei pp. 13-15 pl. ii figs. 3, 8-11, tarri pp. 9-13 pl. ii figs. 4, 5, 12, 13, Strimple (3); P. atoka sp. nov. Carboniferous Oklahoma, U.S., Strimple & Blythe pp. 25-29 pl. iii figs. 4-9 text-fig. 2; P. tarri, Strimple (1) pp. 123-124 pl. ii figs. 7, 10, 13.

†Platycrinites sp., Ivanova p. 134 pl. xviii fig. 7.

† Rhipidocrinus sp. cf. R. perloricatus, Breimer pp. 248-260 text-figs. 1-4 pl. i figs. 1, 2.

†Saccocoma, Lefeld & Radwanski pp. 593-610 text-fig. 1 pls. xxxviii-xli.

†Synbathocrinus melba, Strimple (1) pp. 121-123 text-figs. 1, 2 pl. i figs. 12, 13.

†Synerocrinus incurvus, Ivanova p. 134 pl. xi fig. 3.

† Ulocrinus buttsi, Cronoble pp. 96-99 pl.

Crinoidea A pp. 826–827 pl. xcviii figs. 53–55, B p. 827 pl. xcix fig. 55, Lochman & Hu.

†Crinoids, "intermediate" forms, Regnéll (1) pp. 76-77.

#### MACHAERIDEA

Plumulites cfr. primus pp. 266-267 pl. xiii fig. 17, P. sp. nov. p. 267 pl. xiii figs. 18, 19, Ordovician S. Korea, Kobayashi.

Sokolophocoleus nom. nov. for Lophocoleus Ruedemann, homonym of Lophocoleus Butler, 1886 (Lepidoptera); type Pollicipes siluricus Ruedemann, Pope p. 1054.

## **†EOCRINOIDEA**

Eccrinoids, Regnéll (1) p. 72.

#### †PARACRINOIDEA

Paracrinoids, Regnéll (1) pp. 72-73.

## †CYSTOIDEA

Holocystites alternatus, greenvillensis, Regnéll (8) fig. 3 a-c p. 776.

Hydrophorids, Regnéll (1) pp. 73-76, (3) pp. 775-777.

Cystoid, gen. & sp. indet., Ordovician S. Korea, Kobayashi p. 266 pl.xiv fig. 27.

#### †CARPOIDEA

Carpoidea, Gill & Caster pp. 12-15, Regnéll (3) p. 775.

Superorder Astylophora nov. of subclass Homoiosteles, including order Soluta Jackel, 1900, Gill & Caster p. 11.

Heckericystis gen. nov., family Dendrocystitidae for type H. kuckersiana (Hecker), 1940, Gill & Caster pp. 16-17 fig. 3.

Subclass Homoiostelea nov., Gill & Caster p. 11.

Subclass Homostelea nov., Gill & Caster p. 11.

Iowacystidae fam. nov. of order Soluta, for type genus Iowacystis Thomas and Ladd, 1926, Gill & Caster pp. 15, 20-22.

Iowacystis sagittaria, Gill & Caster fig. 5 p. 20.

Order Mitrata diagnosed, Gill & Caster p. 43.

Mitrocystites mitra, Regnéll (3) fig. 2 p. 775.

Suborder Placocystida diagnosed, Gill & Caster pp. 43-44.

. Placocystites forbesianus, Gill & Caster text-fig. 10 p. 45.

Placocystitidae nom. nov. for Placocystidae Caster, 1952, Gill & Caster pp. 44-46.

Rutroclypeidae fam. nov. of order Soluta, for type genus Rutroclypeus Withers, 1933, Gill & Caster pp. 15, 22–30.

Rutroclypeus emended p. 30 with R. victoriae sp. nov. Devonian Australia pp. 34-40 text-fig. 8 pl. ii fig. 1 pl.iii figs. 1-3 pl. iv fig. 1 pl. vii, R (?) withersi sp. nov. Devonian Australia pp. 40-42 pl. v figs. 2-4 pl. vi fig. 1, "R." globulus pp. 42-43 text-fig. 9, R. junori pp. 30-34 pl. i figs. 1-3 pl. ii figs. 2, 3, Gill & Caster.

Superorder Stylophora nov. of subclass Homoiostelea, including orders Cornuta Jackel, 1900 and Mitrata Jackel, 1921, Gill & Caster p. 11.

Syringocrinus with S. paradoxicus, Gill & Caster pp. 17-21 fig. 4.

Victoriacystis gen. nov. family Placocystitidae for type

V. wilkinsi sp. nov. Silurian Victoria, Australia pp. 46-52 text-fig. 11 pl. vii figs. 1, 2 pl. ix figs. 1, 2 pl. x fig. 2, V. aff. wilkinsi pp. 52-54 text-fig. 12A pl. ix fig. 3, another aff. wilkinsi pp. 54-55 text-fig. 12B pl. x figs. 1, 3, Gill & Caster.

#### †EDRIOASTEROIDEA

Edrioasteroids, Regnéll (1) pp. 77-78, (3) pp. 774-775.

Carneyella pilea, Kesling & Mintz pp. 327-336 figs. 3, 4 pls. v-ix (internal structures).

Isorophus cincinnatiensis, Kesling & Mintz pp. 320-327 figs. 1, 2, pls. i-iv (internal structures); I. austini, Regnéll (3) fig. 1 p. 774.

### †BLASTOIDEA

Blastoids, Regnéll (1) pp. 73-76, (3) pp. 777-781.

Pentremites gutschicki, Regnéll (3) fig. 5 p. 779.

Ptychoblastus gen. nov., aff. Cryptoblastus, for type P. pustulosus sp. nov. Carboniferous Missouri, U.S., Fay pp. 1198-1201 fig. 1 A-F.

#### †OPHIOCYSTIOIDEA

Ophiocistioids, Regnéll (1) p. 78.

